One-Pot α -Glycosylation Method Using Appel Agents in *N*,*N*-Dimethylformamide

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ABSTRACT



A concept for the development of practical glycosylation is presented and demonstrated by one-pot α -glycosylation applying Appel agents for 2-O-benzyl-1-OH hexoses in DMF. The reaction, in situ giving the equilibrium of glycosyl bromides and more reactive O-glycoside intermediates, accomplishes a near-quantitative α -glycosylation removing the water molecules.

Glycosylation involves coupling reactions between glycosyl donors [G1-X (X = leaving groups)] and alcohol or sugar acceptors (G2-OH) to construct α - or β -glycosyl linkages (G1-O-G2). Many types of G1-X are available together with the corresponding promoter agents,¹ and they have contributed enormously to the flexible design of artificial glycoconjugates possessing high medicinal potential.^{2,3} In practice, however, chemical glycosylation is still arduous and far from the straightforwardness of biosynthetic and chemo-enzymatic reactions catalyzed by glycosyltransferases.⁴ Along with our continuous study on the design of glycoconjugate polymers, we have searched for practical glycosylation methods from both chemical⁵ and chemo-enzymatic approaches.⁶ In this

paper, we communicate a chemical pathway that enables us to prepare α -glycosides easily without special care for dehydration of the reaction system.

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For mechanistic reasons, the glycosylation between G1-X and G2-OH must be conducted under anhydrous conditions, except for reactions of simple acceptors under Fisher's conditions or with phase-transfer catalysts.⁷ Though the use of molecular sieves under the pressure of drying gas works effectively against the damage of water contamination, moisture-sensitive reactions cause problems for practical syntheses. More seriously, immobilized desiccants are not adaptable to developing technologies of solid-phase⁸ and fluorous9 syntheses.

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Here, let us hypothesize a dehydrative one-pot glycosylation¹⁰ interfaced with a moisture-sweeping pathway as shown in Scheme 1. There, an $M-X_2$ reagent transforms agents in CH₂Cl₂ yields α -glycosyl bromide α -**2a** (Scheme 2).¹⁴ In this reaction, a hemiacetal 1-OH is trapped in the





1-hydroxyl donor G1-OH to G1-X trapping 1-OH in the form of M=O. Even if contamination by water molecules decomposes G1-X to G1-OH, the excess agent recovers the active donor. Alternatively, the agent may capture every H-OH in a similar way before the decomposition. In both cases, the M-X₂ agent constructs a reaction system of dehydration, donor generation, and regeneration, which is ready for onepot glycosylation with G2-OH. It should be noted that the overall pathway assumed here shows an analogy to conjugate reactions in biosynthetic cascades and the cofactor regeneration system developed by Wong et al.¹¹ for mammalian transferases.

To realize our hypothetical pathway, the choice of $M-X_2$ agents is a key investigative issue. The reagent should be more reactive to G1-OH and H-OH than to acceptor G2-OH. The reagent itself, M=O, and other possible secondary products should not affect the glycosylation. Among several candidates of $M-X_2$, we have evaluated the potential of a reagent combination of triphenylphosphine (Ph₃P) and carbon tetrabromide (CBr₄) known as Appel agents.¹²

In our previous study,¹³ we found that the reaction of 6-*O*-acetyl-2,3,4-tri-*O*-benzyl-D-glucopyranose **1a** with Appel



form of $Ph_3P=O$, showing a feature matching the conversion of $M-X_2$ to M=O. In the present study, we investigated an optimal access to one-pot glycosylation as well as possible dehydrating pathways.

First, we addressed a halide ion-catalytic α -glycosylation pioneered by Lemieux and co-workers.¹⁵ After a mixture of **1a** and Appel agents was stirred in CH₂Cl₂ for 2.5 h, cholesterol **3a**, *N*,*N*-tetramethylurea,¹⁶ and Et₄NBr were added successively to the mixture. Excitingly, we were able to obtain α -glucosylated cholesterol in high yields and α -selectivity (entry 1, Table 1), though no molecular sieves

Table 1. One-Pot Glycosylation with G1-OH Donors and Appel Agents in Either CH_2Cl_2 or DMF^a

					α/β
entry	G1-OH	R ² -OH	conditions	\mathbf{h}^{b}	(yields, %)
1	1a	3a	Et ₄ NBr+TMU/CH ₂ Cl ₂	53	98:2 (92)
2	1a	3a	TMU /CH ₂ Cl ₂	83	>99:1 ^d (90)
3	1a	3b	TMU/CH ₂ Cl ₂	96	>99:1 ^d (92)
4	1a	3a	DMF	24	95:5 (96)
5	1a	3b	DMF	22	>99:1 ^d (92)
6	1b	3a	TMU/CH ₂ Cl ₂	58	90:10 (95)
7	1b	3a	DMF	23	85:15 (95)
8	1c	3a	DMF	23	94:6 (92)
9	1d	3a	DMF	23	82:18 (96)
10	1e	3a	DMF	19	86:14 (95)

^{*a*} All reactions were carried out at room temperature with donors (0.4 mmol), acceptors (1.2 mmol), and a mixture of CBr₄ and Ph₃P (1.2 mmol for each) in given solvent. ^{*b*} Reaction times (h) until every donor was consumed. ^{*c*} Isolated yields based on the amount of 2-*O*-benzyl-1-OH donors. ^{*d*} No β -isomer was detectable in the ¹H NMR spectra of products.

and drying gas were applied. Second, we tried to further simplify the reaction conditions. Entry 2 shows that the α -glycosylation proceeded even in the absence of tetraethyl-ammonium bromide, though it took a prolong time to completion. Entry 3 shows that the simple reaction is valid

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⁽¹⁴⁾ An analogous reaction was reported by Khatuntseva et al.^{12d} who applied the Appel agents for 1-bromination of L-fucose derivatives and onepot glycosylation in the presence of HgBr₂, Hg(CN)₂, and molecular sieves.

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also for a sugar acceptor **3b** with a hindered 3-OH, proving its potential for oligosaccharide synthesis.

When the overall reaction was carried out in DMF, glycosylation proceeded even in the absence of tetraethylammonium bromide, the one-pot α -glycosylation could be additionally simplified and facilitated (entries 4 and 5). In this case, *N*,*N*-tetramethylurea was replaced with the solvent itself. Entries 6–10 show that every D-gluco- (**1a**, **1b**), D-galacto- (**1c**, **1d**), and L-fucosyl **1e** donor gave the corresponding α -glycosides efficiently. When the α -selectivity of **1a** (entry 3–5) and **1c** (entry 8) with a 6-*O*-acyl group is compared with that of **1b** and **1d** with a 6-*O*-benzyl group, we can see the effect of the 6-*O*-acyl group to promote α -selectivity (Scheme 3).¹⁷



To search for the underlying dehydration pathways, we undertook ¹H NMR study for the reaction of **1a** and Appel agents. After the 1-bromination was carried out for **1a** in CH₂Cl₂, the mixture was concentrated and dissolved again in either CDCl₃, DMF- d_7 , or ca. 1% D₂O–DMF- d_7 solvent. The ¹H NMR spectrum of the CDCl₃ solution enabled us to confirm that the reaction gave α -**2a** exclusively (H-1, δ 6.40 ppm, doublet, $J_{1,2} = 4.0$ Hz, Figure 1a). On the other hand, the spectrum of the DMF- d_7 solution showed a feature different from that of α -**2a**. The H-1 signals shifted to a lower field, while it kept the vicinal coupling constants character-



Figure 1. ¹H NMR spectra (500 MHz) of a mixture of **2a** and Appel agent dissolved again in $CDCl_3$ (a) and 1% D_2O-DMF solvent (b). [Spectrum c was recorded after Appel agents (9 mol equiv for **1a**) were added to the ca. 1% D_2O-DMF solution. Asterisk (*) notes signals of unidentified nonsugar components.]

istic for α -D-glucopyranose with ${}^{4}C_{1}$ -ring conformation (H-1, δ 7.08 ppm, d, $J_{1,2} = 4.0$ Hz: H-2, δ 3.68 ppm, dd, $J_{2,3} = 10.0$ Hz). This indicates that the glycosyl bromide α -**2a** is transformed to an alternative α -glycosyl species α -**2a**-**X** in the DMF- d_{7} solution. Another experiment reacting **1a** and Appel agents in an NMR glass tube allowed us to determine that α -**2a**-**X** is derived in DMF solvent but not in CH₂Cl₂. In Figure 1b, it is shown that α -**2a**-**X** is unstable in an aqueous DMF- d_{7} solvent and returns to a mixture of α -**2a** and **1a** in a reversible manner. This also shows that the new species is more sensitive to water than α -**2a**. Figure 1c shows that an additional introduction of the Appel agents regenerates the labile species, removing every water molecule.

In these NMR experiments, we observed some intriguing phenomena as described below. (1) The reagents begin the conversion of **1a** to α -**2a**-**X** after they dry up the solvent system. (2) The capacity of water absorption is obviously larger than what we expect for the conversion of Ph₃P to Ph₃P=O. It is obvious that CBr₄ is also used for the dehydration of DMF probably to yield CO₂ or CO₃²⁻ in a final form. (3) The ¹H-signals of α -**2a** are hardly detectable on the way leading to α -**2a**-**X**, while they appear upon the addition of D₂O to α -**2a**-**X** in the DMF solution leading back to **1a** as can be seen in Figure 1b. (4) Every conversion among the three sugar components, i.e., **1a**, α -**2a**, and α -**2a**-**X**, is reversible and with negligible side reactions affording irreversible sugars.

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The above ¹H NMR study as well as mechanistic considerations allow us to assign the chemical structure of α-2a-X as O-[6-O-acetyl-2,3,4-tri-O-benzyl- α -D-glucopyranosyl]methylene-N,N-dimethylammonium bromide, a kind of Vilsmeier-Haack intermediate.¹⁸ The FAB-mass spectrum of a mixture of α -2a-X and the Appel agents in DMF supported this assignment $[(M + Na)^+ = 650]$. Because of its highly labile property, the spectral assignment of α -2a-X may remain incomplete. However, the formation of such an intermediate can rationalize not only the NMR observations but also the glycosylation reactions accelerated in DMF. Though Lemieux and co-workers¹⁵ reported a similar solvent effect, they did not indicate the occurrence of such DMFglycosyl adducts. Thus, it seems appropriate to propose a novel α -glycosylation pathway where the reactive O-glycoside is in situ formed and associated (Scheme 4). We



speculate that the intermediate may accelerate the formation of β -glycosyl bromide leading to α -selective glycosylation. It is also probable that the glycosyl imidate like α -**2a**-**X** is derived not only from glycosyl bromide but also directly from 1-OH sugar in the reaction with a Vilsmeier reagent produced in the Appel agents-DMF system. It is obvious in any case that the Appel agents play multiple roles and thus allow a simple α -glycosylation.

In conclusion, we have demonstrated a general strategy to develop practical glycosylation, in which the choice of M-X₂ type reagent for G1-OH type donors is a key investigative issue. In this paper, we have described the convenient use of Appel agents in DMF and thereby proposed a one-pot α -glycosylation method. The glycosylation via a glycosyl imidate looks similar to the Otrichloroacetimidate method established by Schmidt.^{1b} More exactly, the dehydrative method using 2-O-benzyl-1-OH donors is closer to one-pot dehydrative glycosylation of Gin and co-workers,10 Koto et al.,19 Leroux and Perlin,20 and Pavia et al.²¹ Our method is, however, distinct from these methods in that the reaction takes place in the universal. nonhalogenated, but hygroscopic solvent DMF without special care for dehydration.²² This property will have implications in scaled-up, solid-phase, and other glycosylation processes.

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Supporting Information Available: ¹H NMR and FAB-MS data of glycosyl products and ¹H NMR spectra supporting reversible conversion among **1a**, α -**2a**, and α -**2a**-**X**. This material is available free of charge via the Internet at http://pubs.acs.org.

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⁽²²⁾ A General Protocol for One-Pot α -Glycosylation in DMF. Overall, reactions are conductible at ambient temperature (15–20 °C) in a glass vessel closed with a septum cap. No molecular sieves or drying gas is required as long as the vessel is dried at 120 °C in an oven prior to the use. Typical procedure is as follows: A 2-O-benzylated sugar G1-OH in DMF (molecular sieves 4A inclusive, 99.8% grade, water content <0.1%) is treated first with a mixture of Ph₃P (3 mol equiv) and CBr₄ (3 mol equiv) until G1-OH sugar is consumed completely (observed as glycosyl bromide on silica gel TLC, *n*-hexane/ethyl acetate = 5:1) (2–3 h) and then with an acceptor alcohol (2–3 molar equiv). The reaction is stirred at room temperature until the donor is consumed (8–25 h). The reaction mixture is diluted in toluene, washed with saturated NaHCO₃ and water, concentrated, and subjected to purification on silica gel column.